

RNAi technologies to control pests and pathogens in canola

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Two major threats to our canola

Sclerotinia – a fungal pathogen



Flea beetles – a pest insect



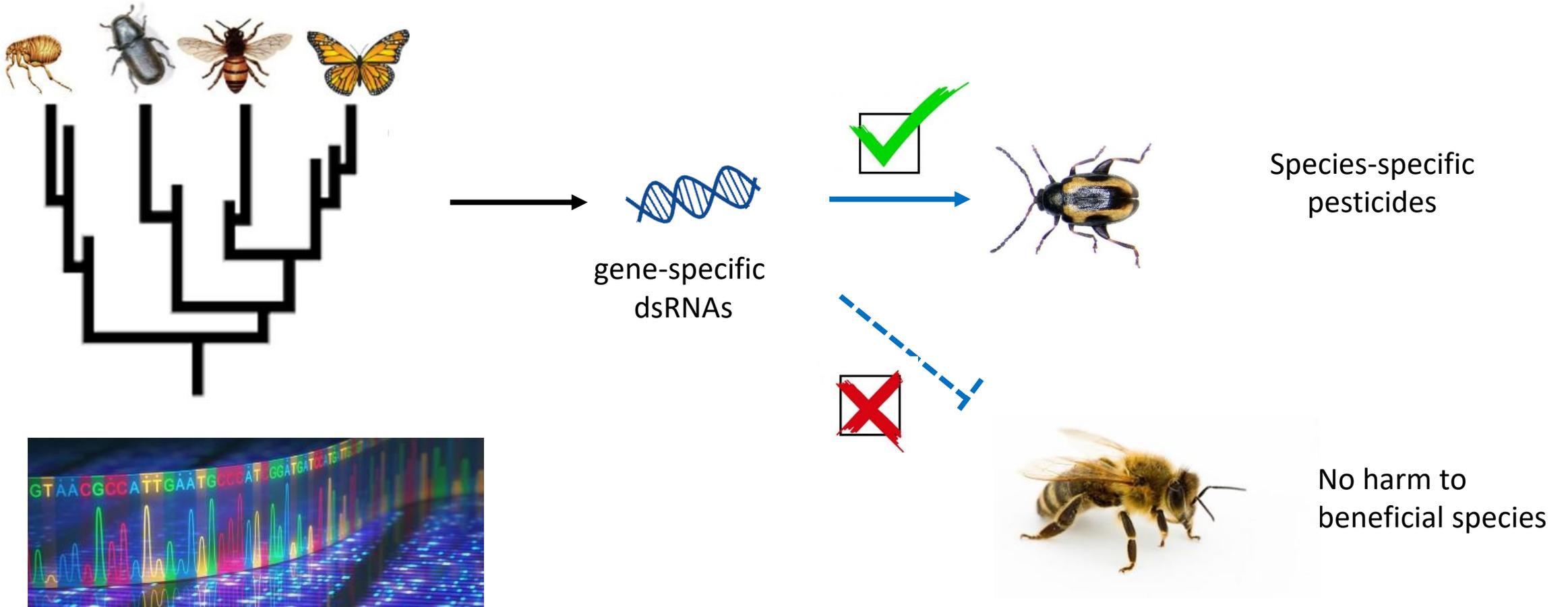
The current approach to protecting canola - chemical pesticides

Two issues of concern about pesticides:

1. Increasing resistance
2. Off-target effects

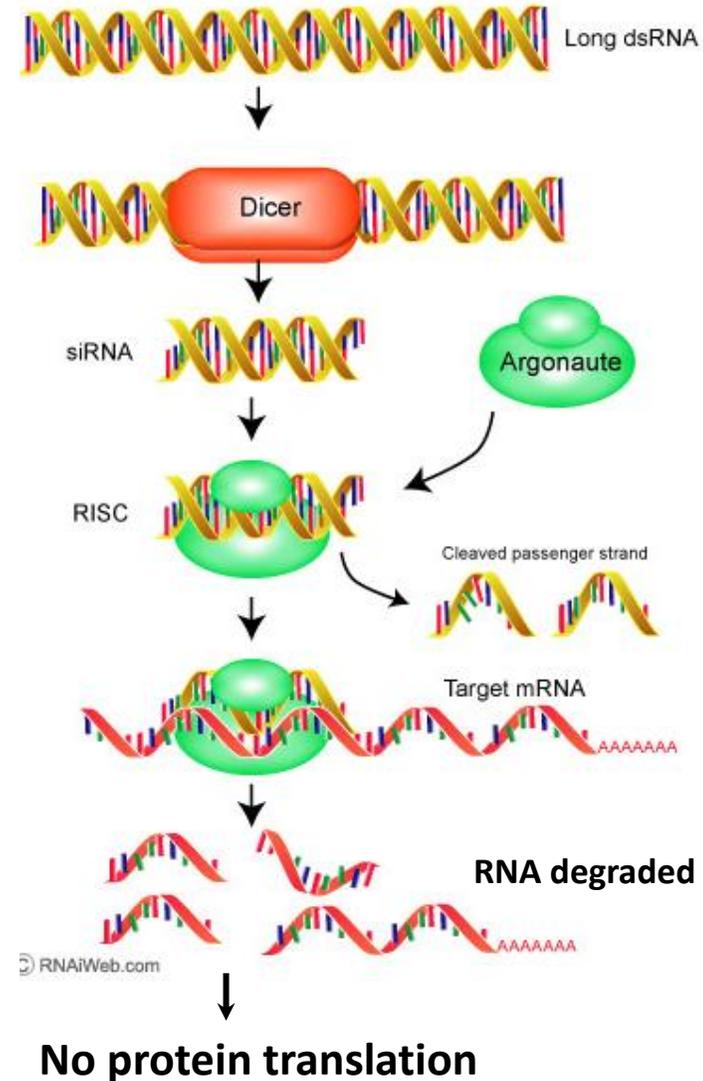


Species-specific dsRNA pesticides/fungicides



RNA-based pesticides – using RNA interference

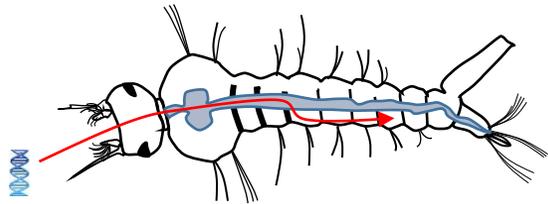
- RNAi = gene silencing using double-stranded RNA
- Useful tool for creating loss-of-function mutants
- Potential to kill pest insects?



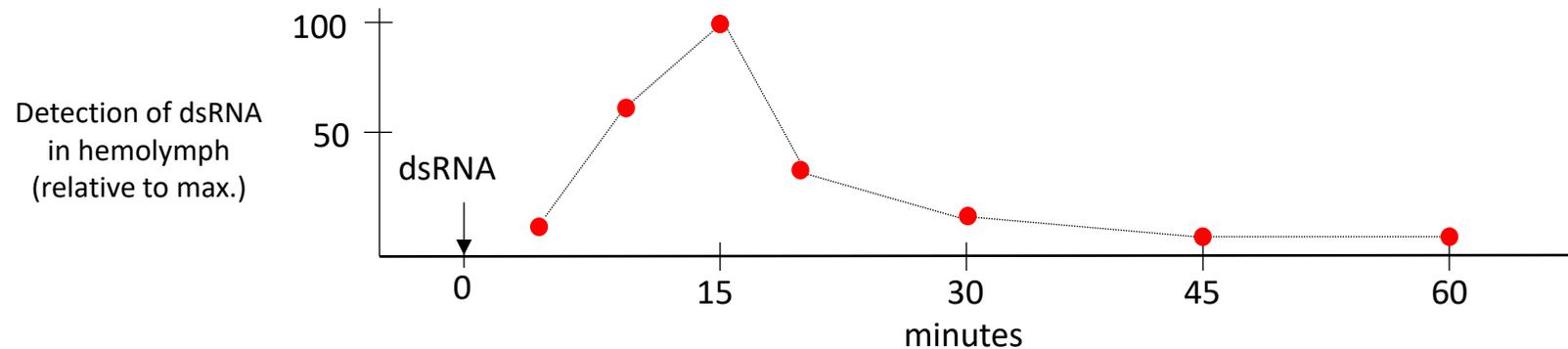
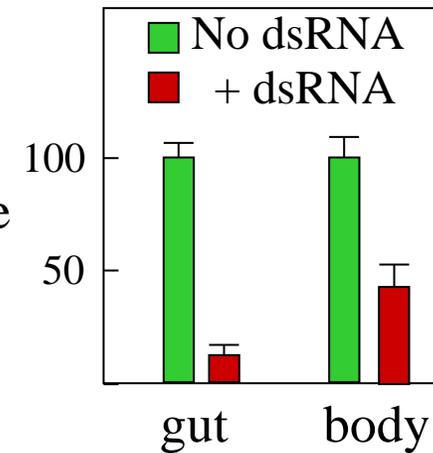
Feeding dsRNA to insects

- Ingested dsRNA can silence genes in the insect gut
- The dsRNA can spread and silence genes in other tissues (systemic RNAi)

mosquito larvae fed dsRNA



tubulin gene expression



Flea beetles

- Pest species in Manitoba:
 - *Phyllotreta cruciferae* (crucifer flea beetle)
 - *Phyllotreta striolata* (striped flea beetle)
- Feed on cruciferous plants
 - including canola



P. cruciferae



P. striolata



Developing RNAi pesticides vs flea beetles



Field collections



Lab colonies

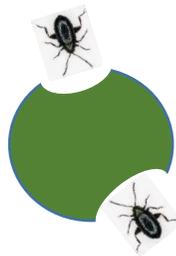


RNA sequencing

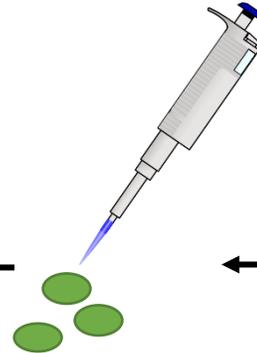
Assess mRNA knockdown

Assess mortality

Assess leaf damage



Feeding bioassays



DsRNA on leaf discs

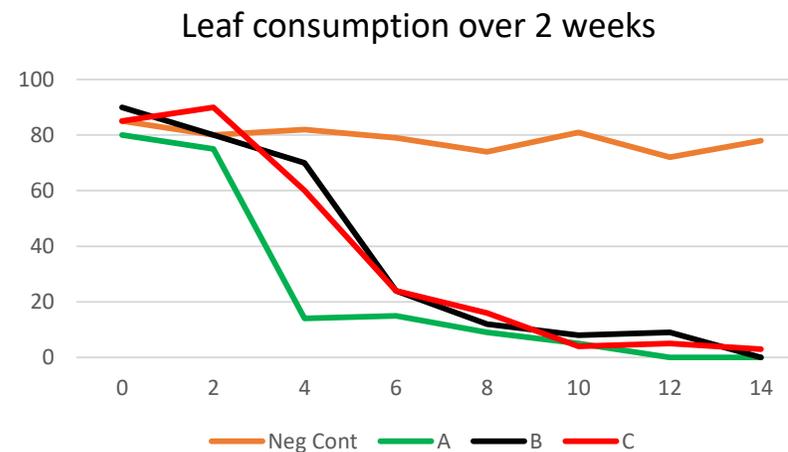
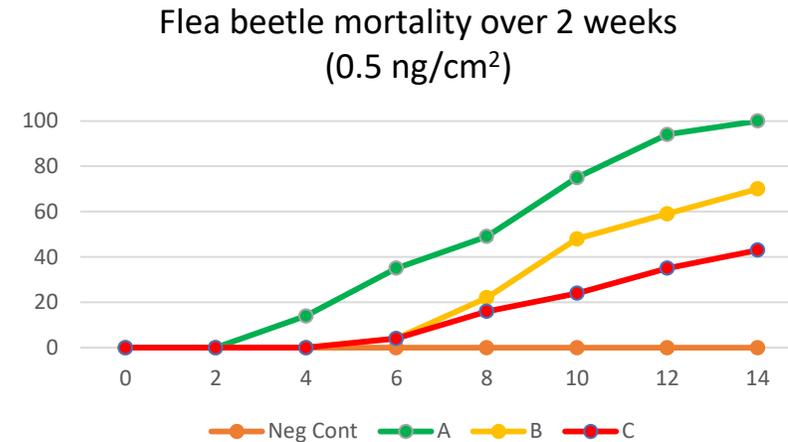


Identify flea beetle-specific gene sequences

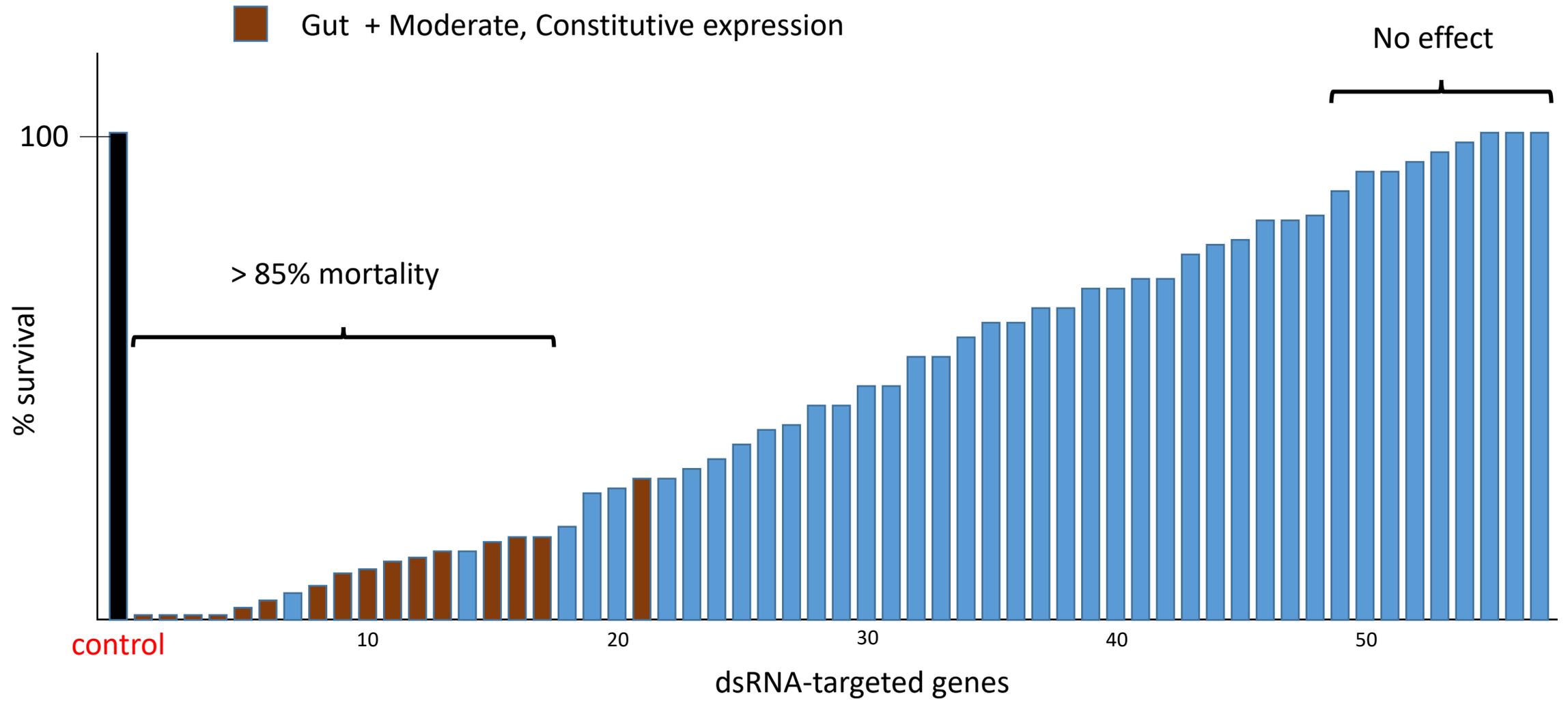


Control of flea beetles on canola

- Transcriptomic data identified >100 target genes tested
- Topical treatment of leaves with dsRNA kills flea beetles
 - But slower than neurotoxic chemicals
- Leaf damage was reduced
 - Insects stopped feeding after 4 days



Identifying ideal RNAi insecticide target genes



Species-specific RNA insecticides

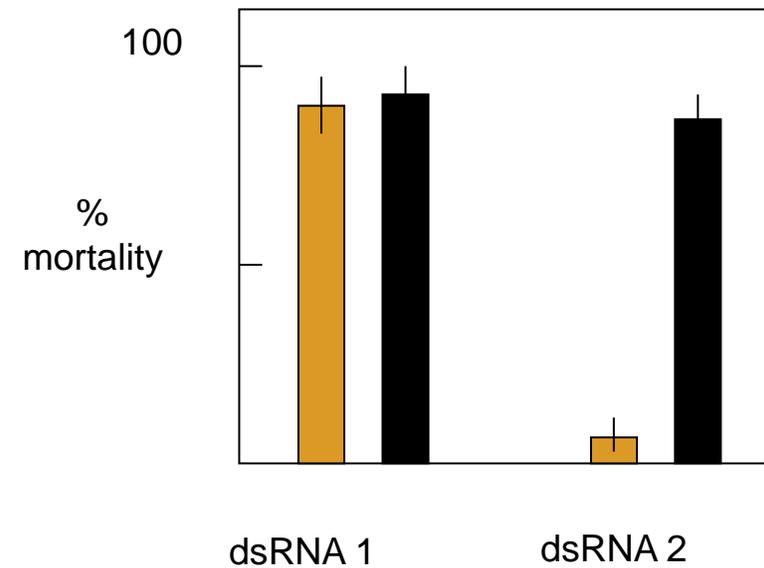
Flea beetle RNAi insecticides



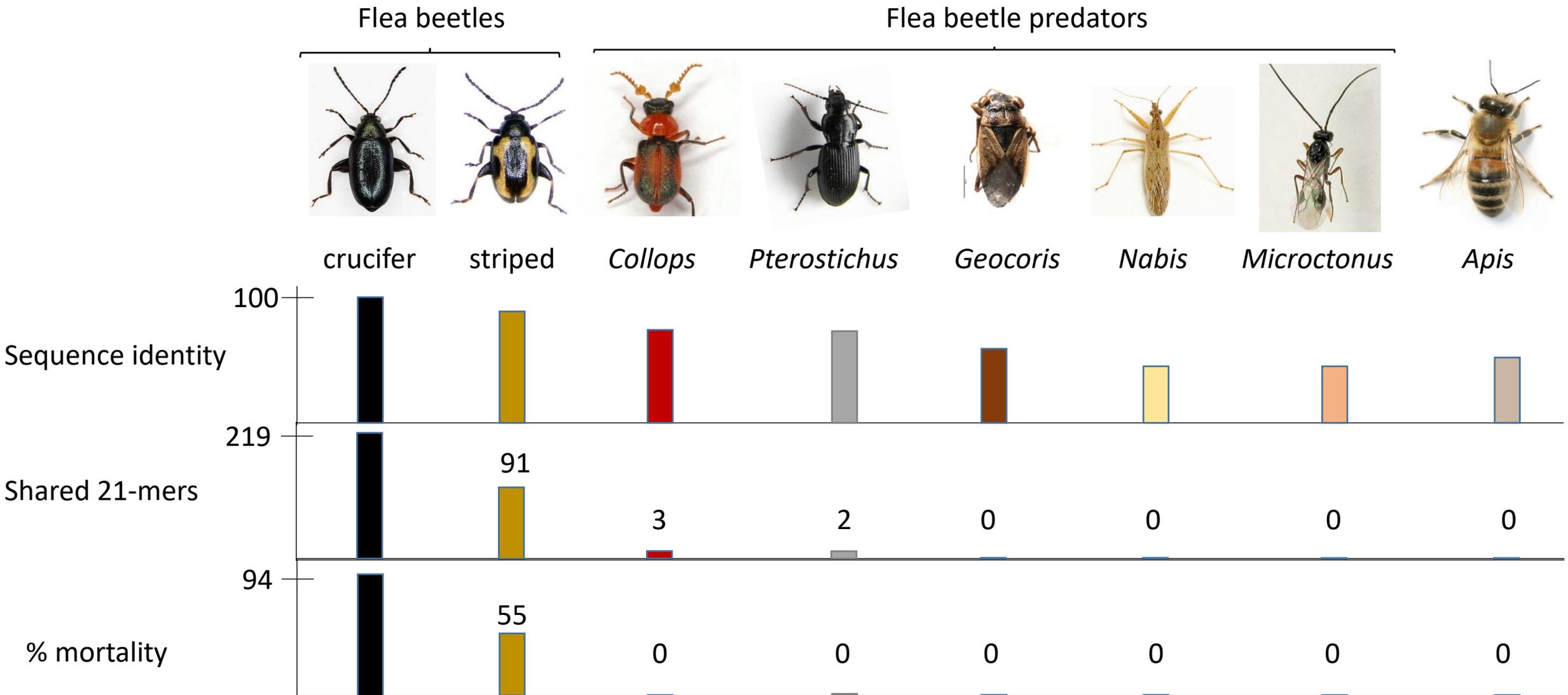
Striped flea beetles



Crucifer flea beetles

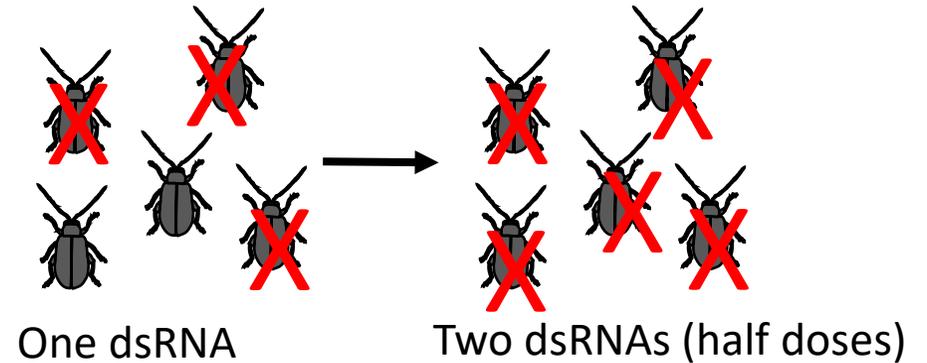


Specificity of a dsRNA pesticide in the canola agrosystem



Can we improve RNAi efficiency?

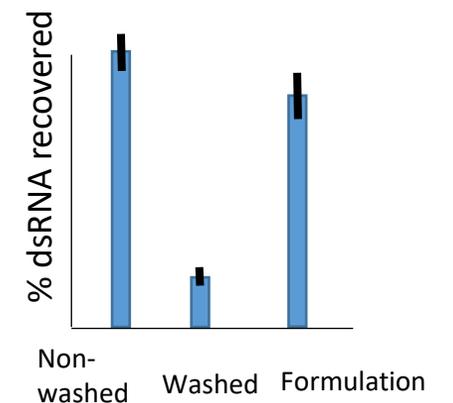
- Easy to combine different dsRNAs
 - Improved percentage kill up to 100%



- Improved RNA stability
 - Reduced dsRNA by >50%



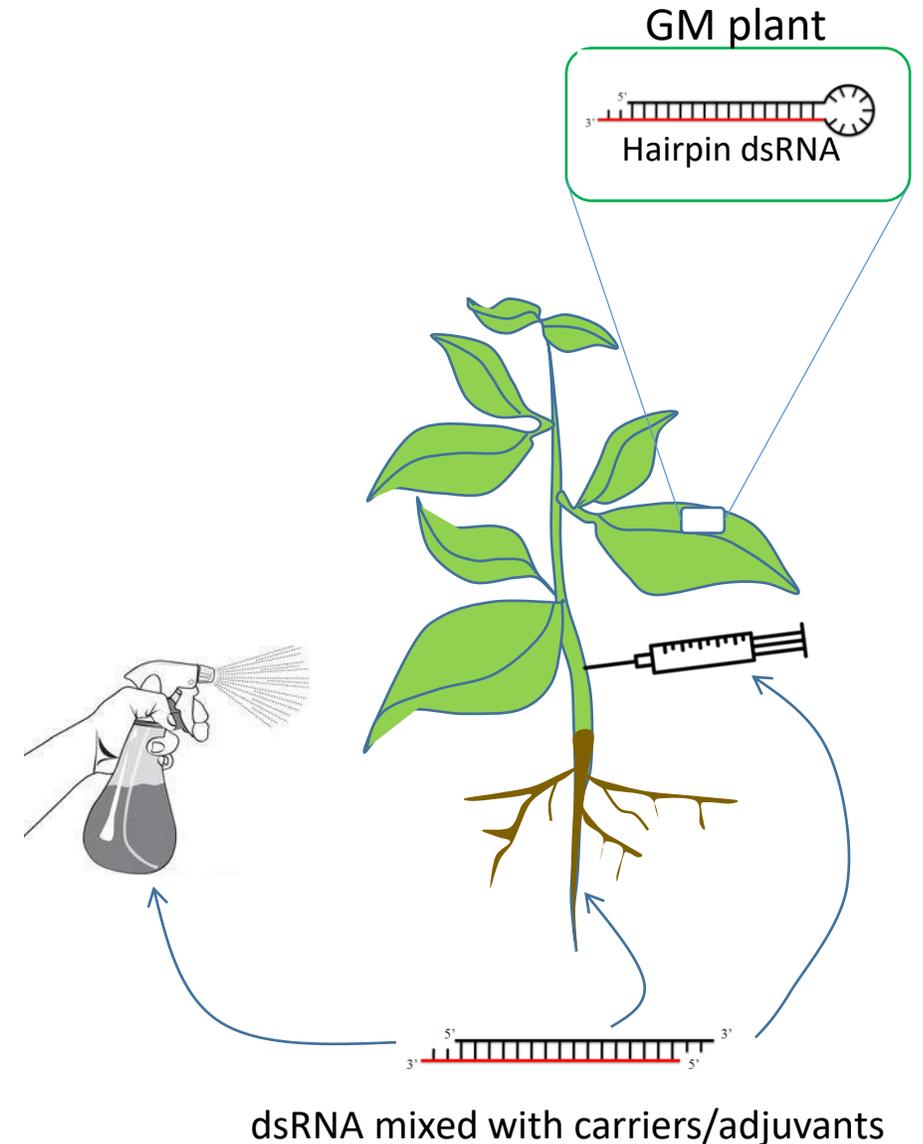
- Improved formulations
 - Did not wash off, remained active



RNAi-based crop protection

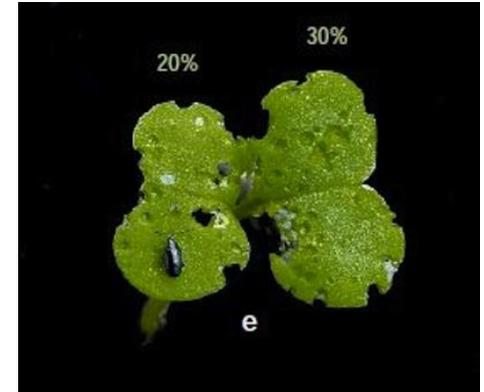
Different dsRNA delivery approaches:
(not just canola!)

- In-planta (transgenic approach)
- Foliar sprays
- Root delivery
- Stem injections

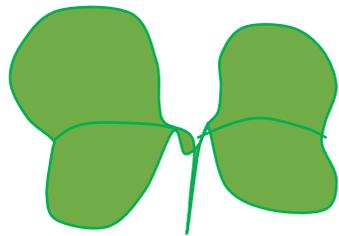


Comparing transgenic canola vs foliar dsRNA pesticides

- Canola and Arabidopsis transformed with snf7 targeting *Phyllotreta* spp
- Comparing flea beetle damage

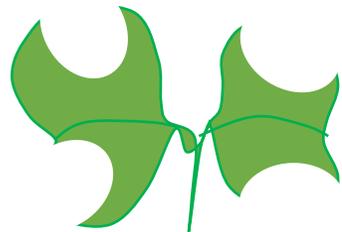


No beetle control



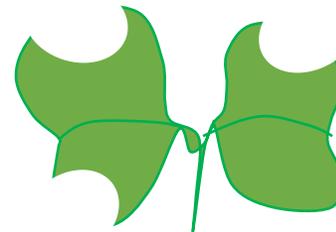
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No dsRNA control



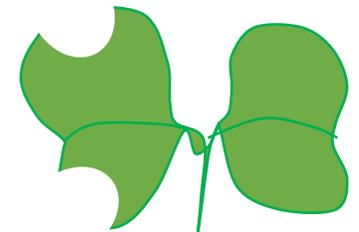
64 ± 14

Transgenic –hairpin-snf7



24 ± 4

Foliar hairpin – snf7



18 ± 5

% leaf consumption:

Beetle mortality:

N/A

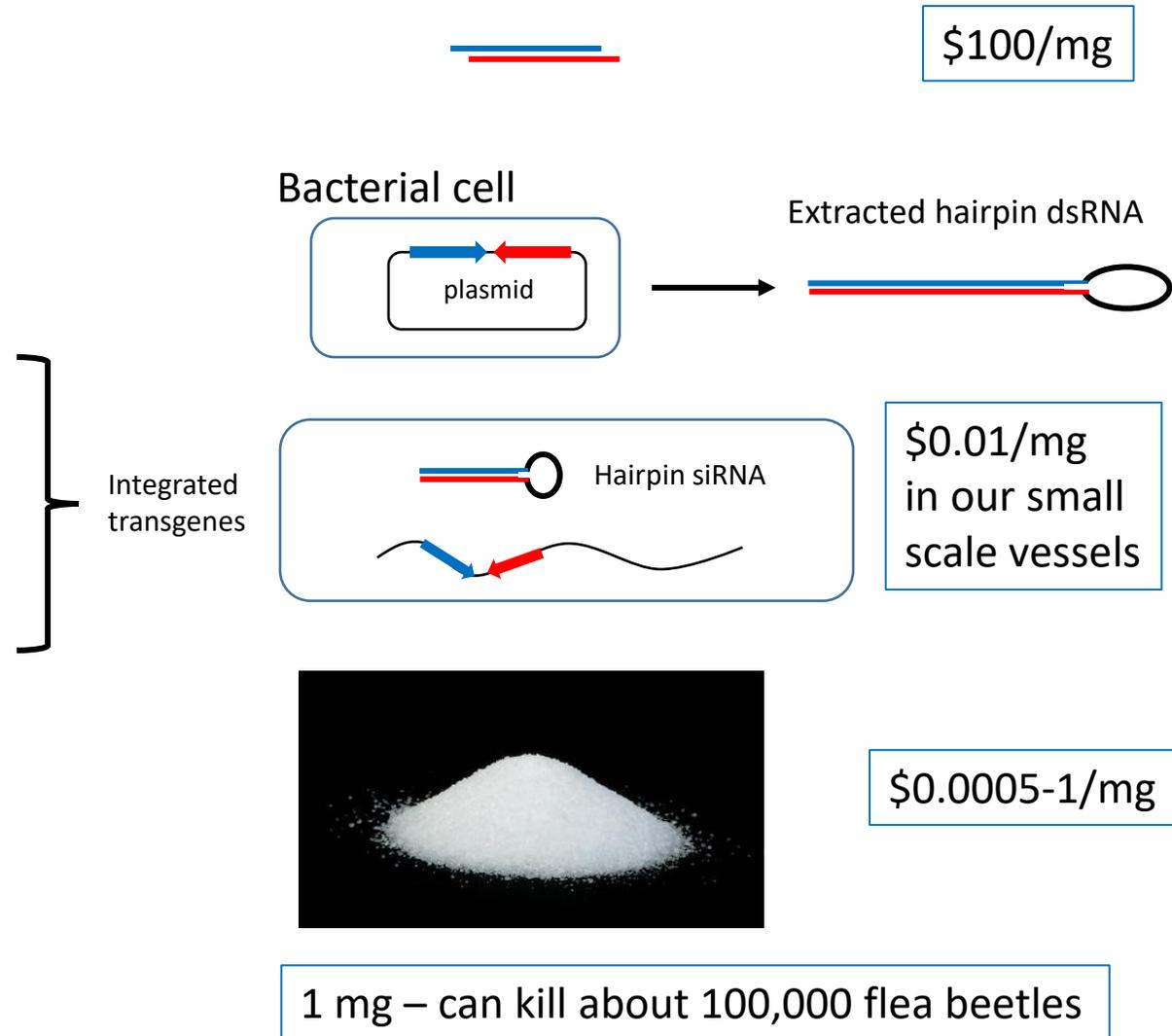
3 ± 2

38 ± 9

68 ± 12

Is the technology affordable?

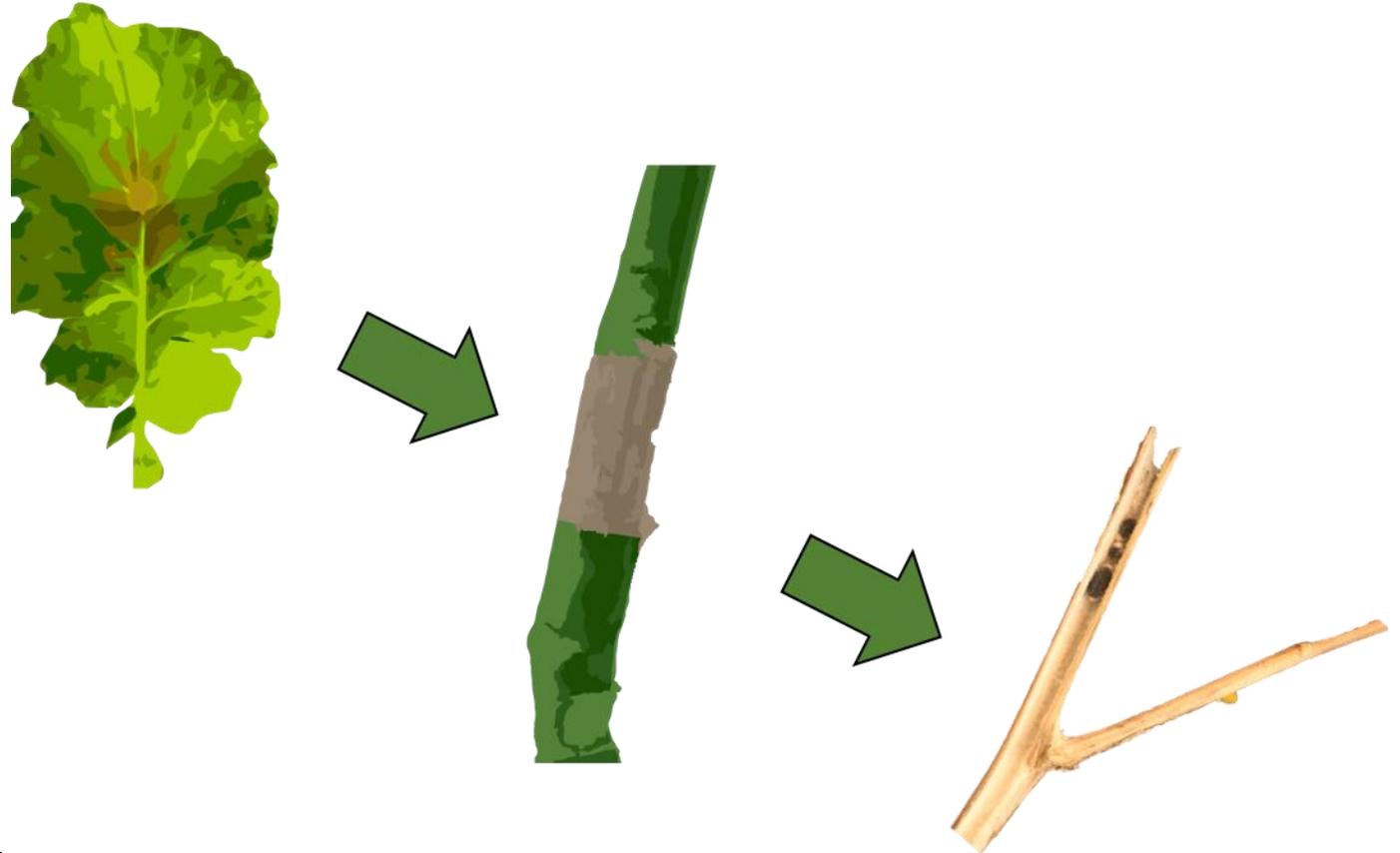
- In vitro synthesis
- Bacterial production
- Yeast production
- Algae
- Commercial producers



Anti-fungal dsRNA: target *Sclerotinia*

- *Sclerotinia sclerotiorum*

- A major cause of yield loss
- Over 450 host plants
- 90% of Canadian canola fields show infection
- Causes millions in lost yields annually
- Life cycle makes control difficult with conventional methods



RNA sequencing to identify *Sclerotinia* infection genes



Susceptible-24h

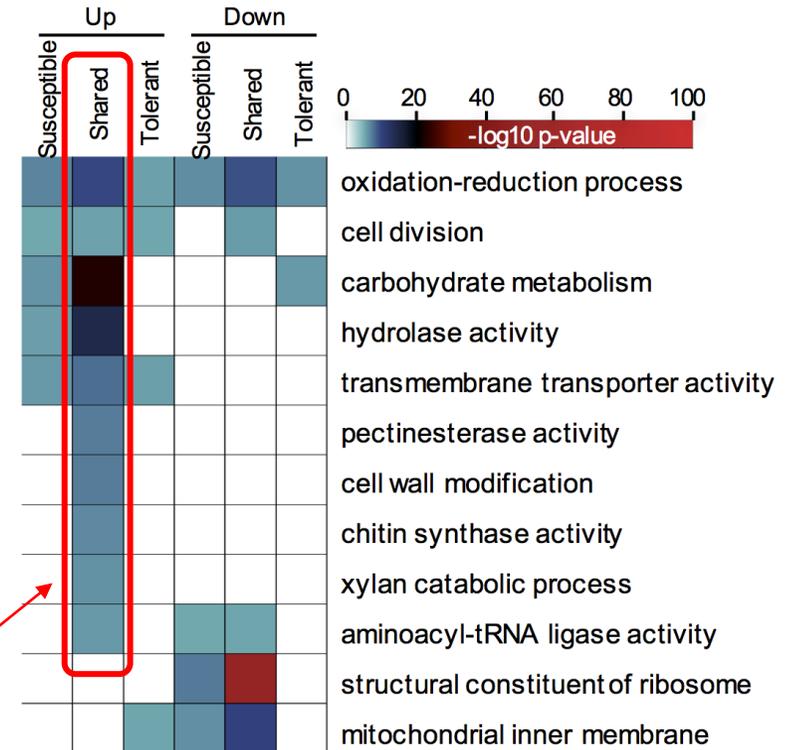


Tolerant-24h

1. Infect different strains of canola



2. RNA sequencing



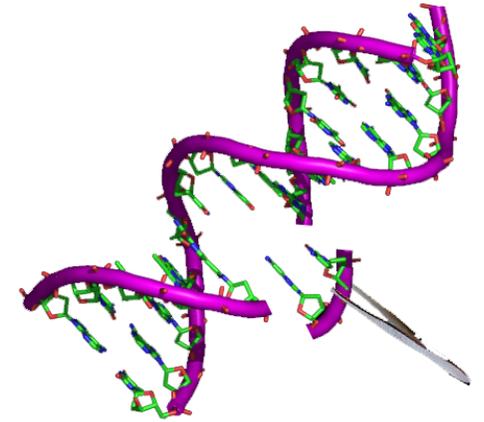
3. Identify genes up-regulated during infection

Two methods of protection using RNAi

1. Transgenic plants expressing dsRNA

Advantage: Entire plant protected all the time

Disadvantage: Public perception and regulatory delays



2. Foliar dsRNA sprays

Advantage: Easily transferrable to more crops

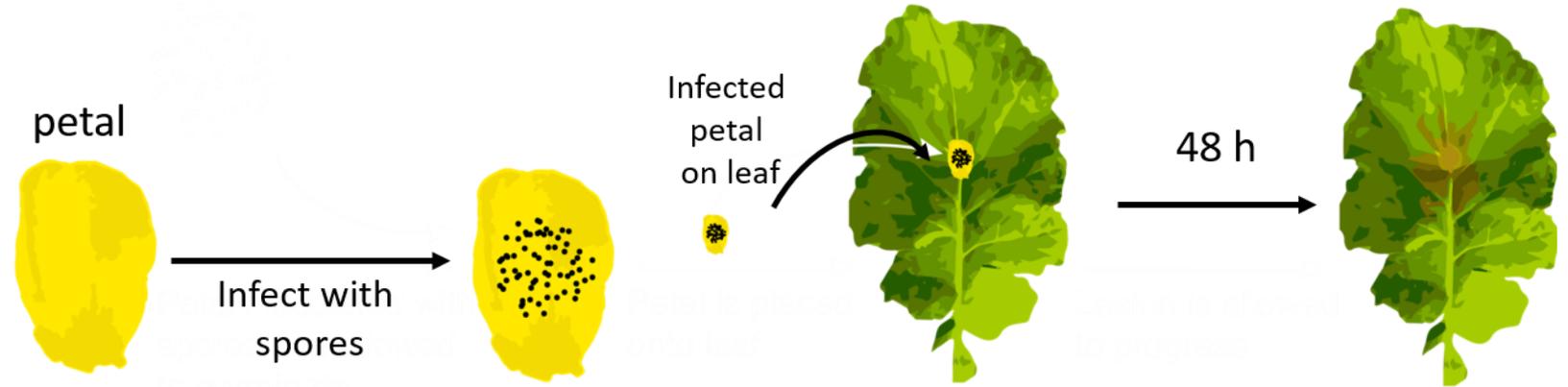
Disadvantage: Can require multiple treatments (\$)



Two infection assays

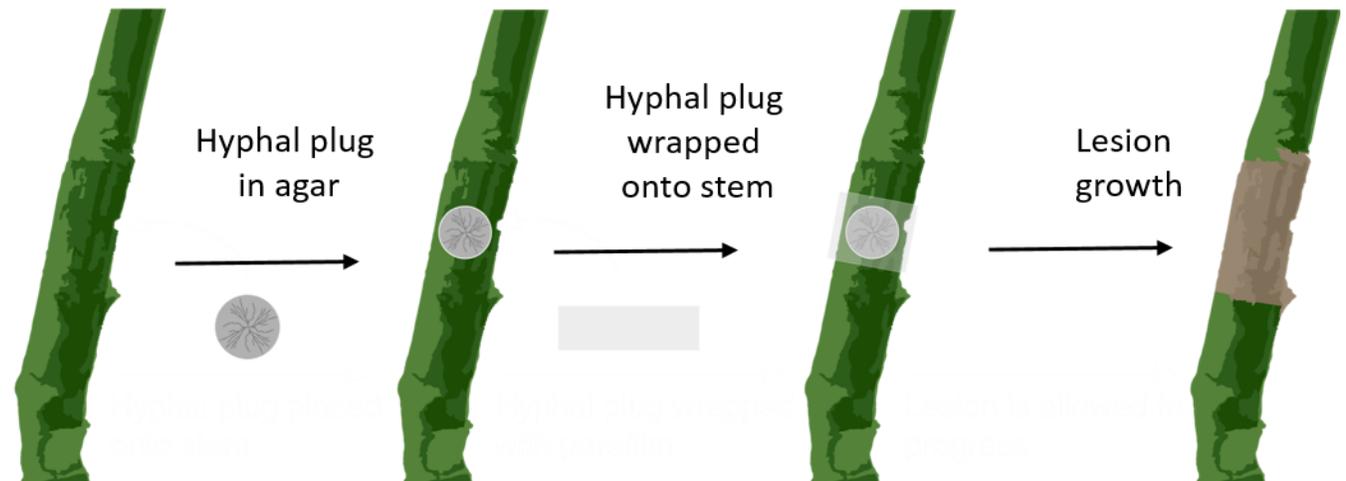
Leaf lesions

- Used mostly for testing foliar dsRNA applications



Stem lesions

- Used mostly for testing transgenic plants

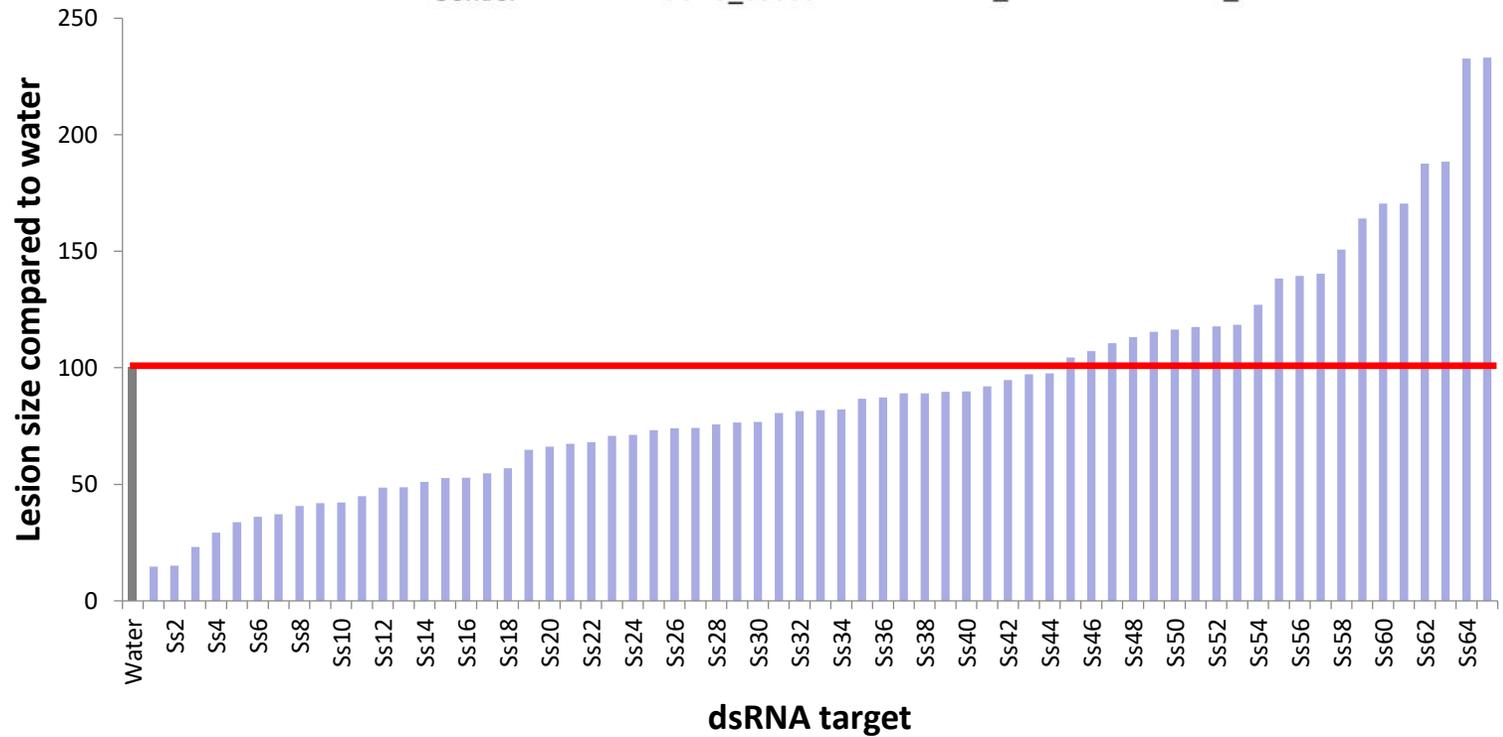


DsRNA screening

Leaf lesion assays

>150 dsRNAs screened

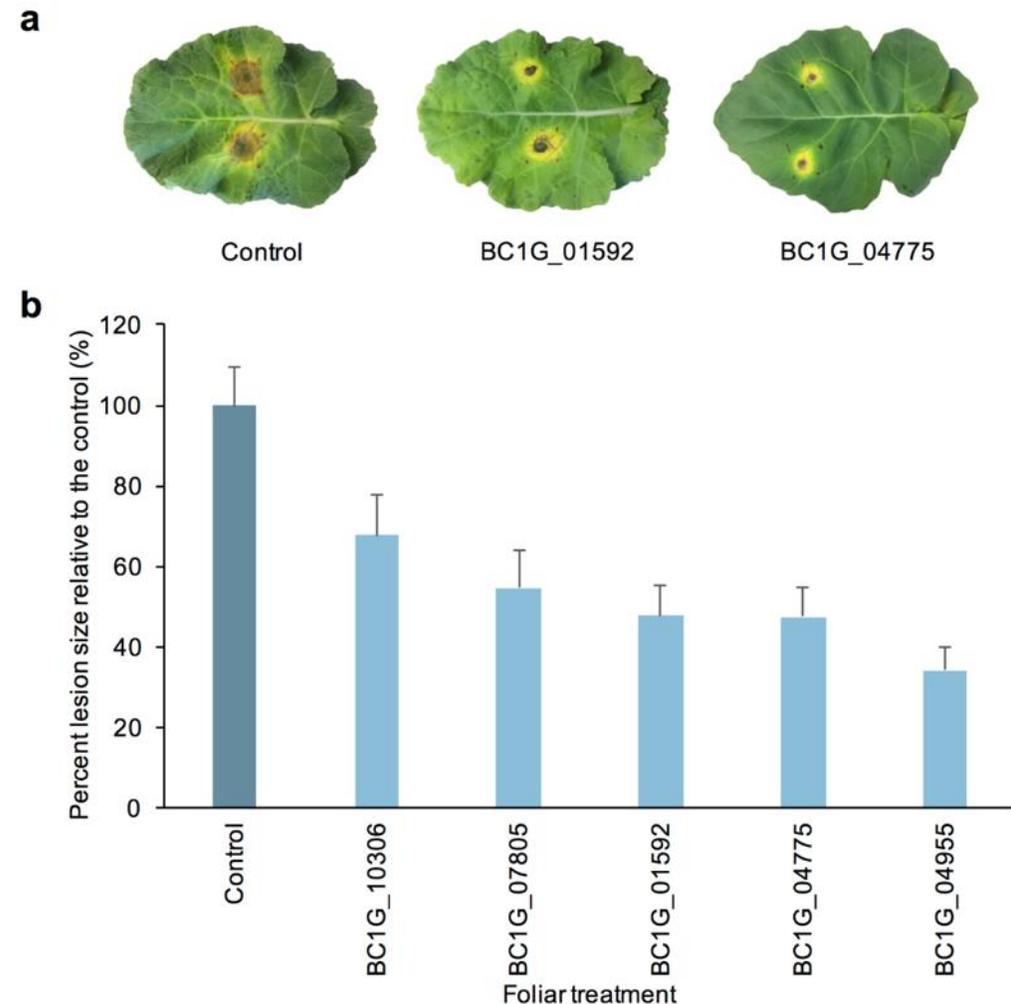
>20 dsRNAs strongly inhibited
Sclerotinia infections



Can RNAi be used against other fungal pathogens?

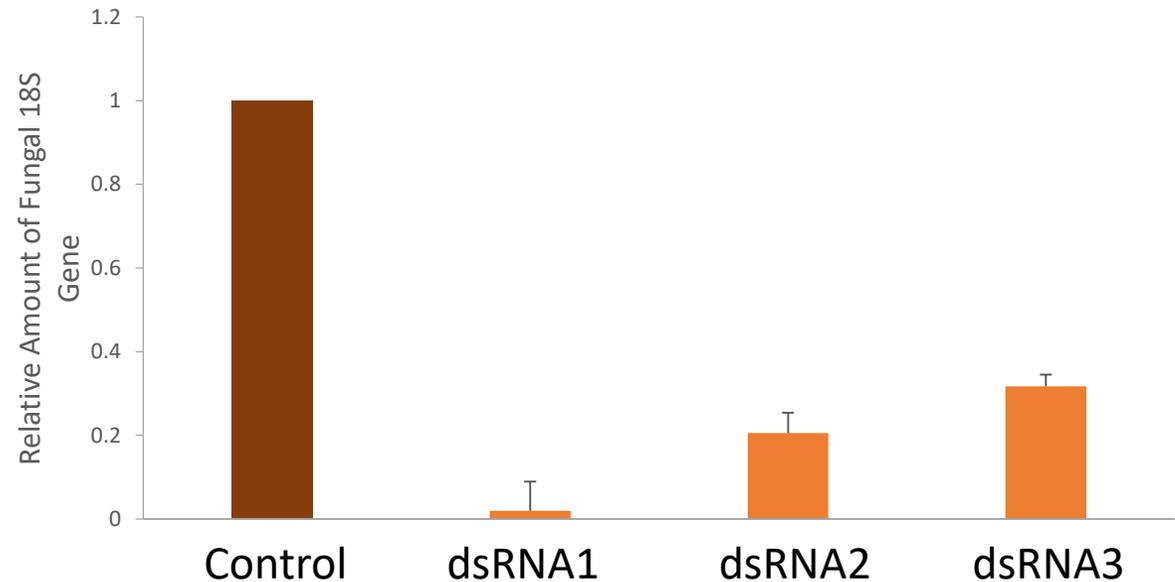
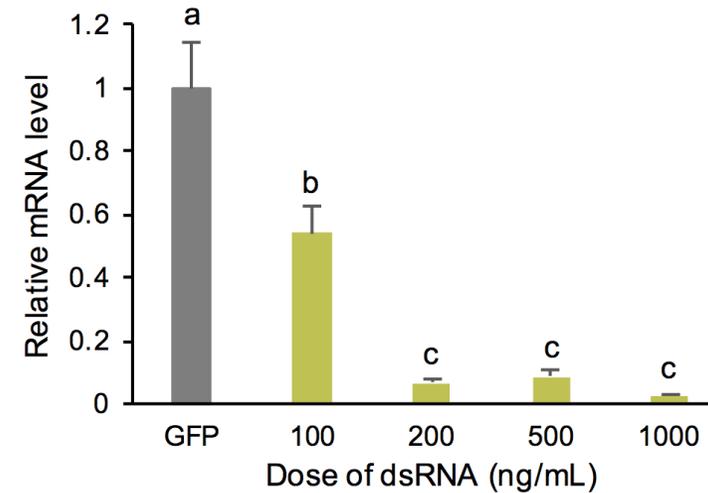
The technology translates to other fungal pathogens

Some of the same genes targeted in *Sclerotinia* can be effective targets against *Botrytis*

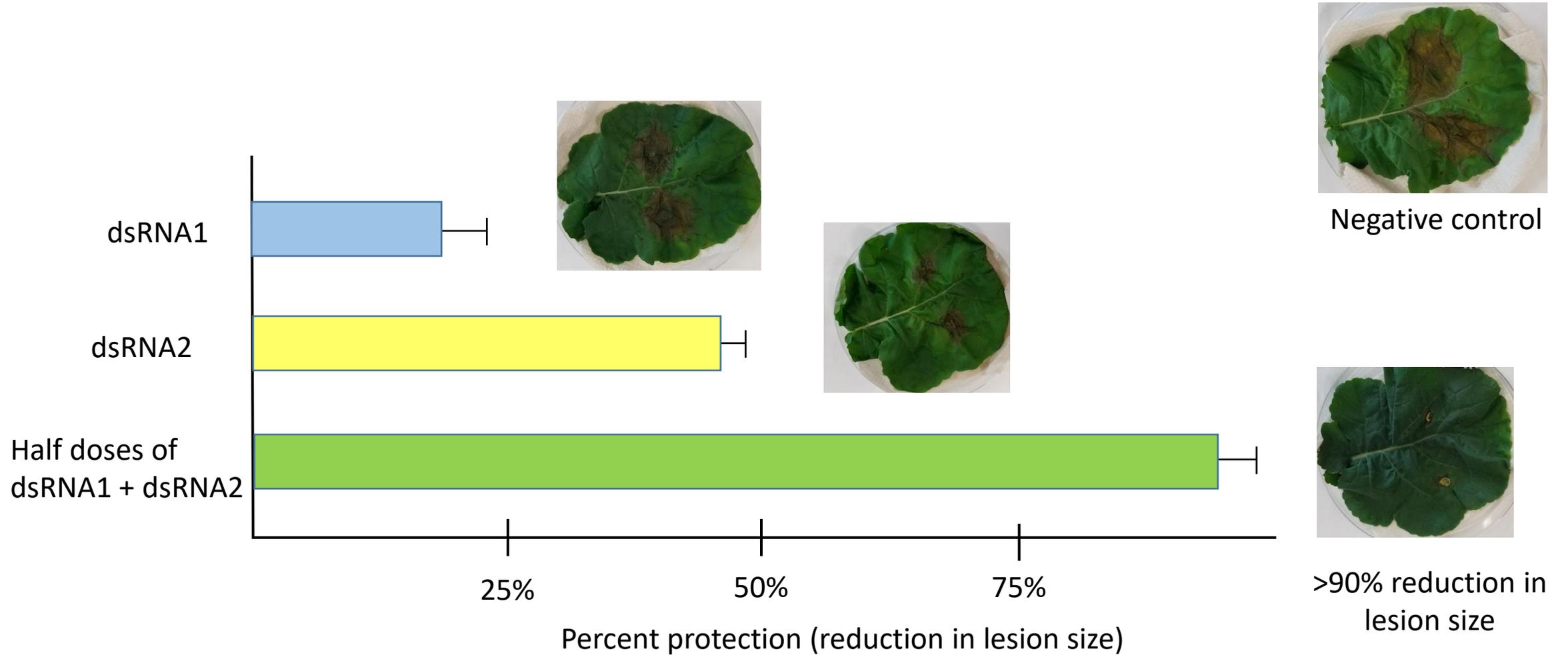


RNAi reduces fungal transcripts and fungal load

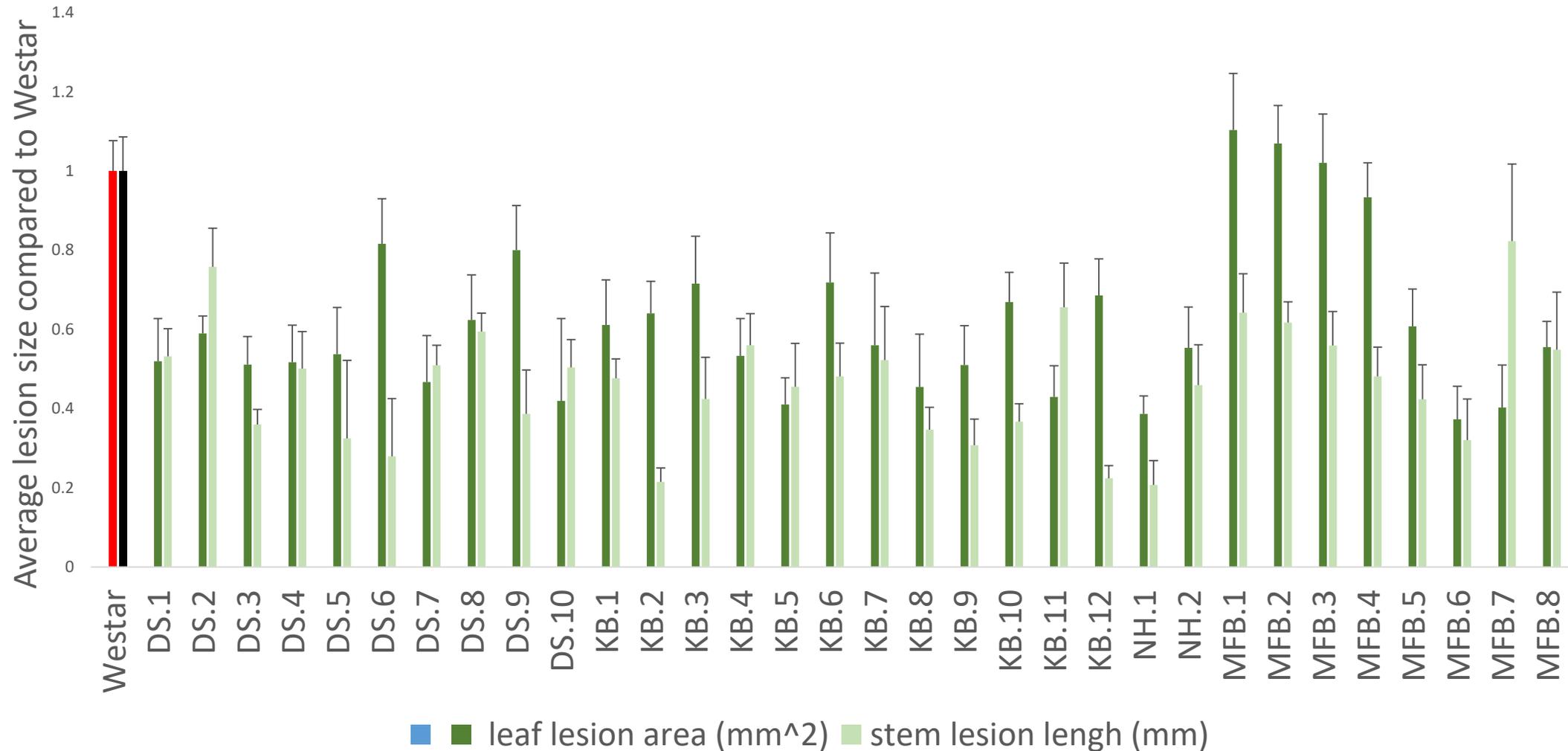
- qRT-PCR confirms that dsRNA is reducing the targeted mRNAs
- qRT-PCR confirms that the amount of fungus in the lesion region is reduced



Can combining dsRNAs improve protection?



Are transgenic plants protected by RNAi?



What does protection look like?

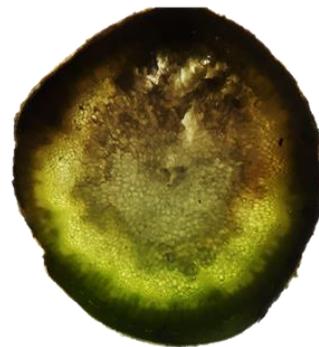
Westar

Transgenic

3 dpi



3 dpi



How about whole-plant protection?



Mature canola plants were sprayed with highly concentrated *Sclerotinia* spores in high humidity (1 week)

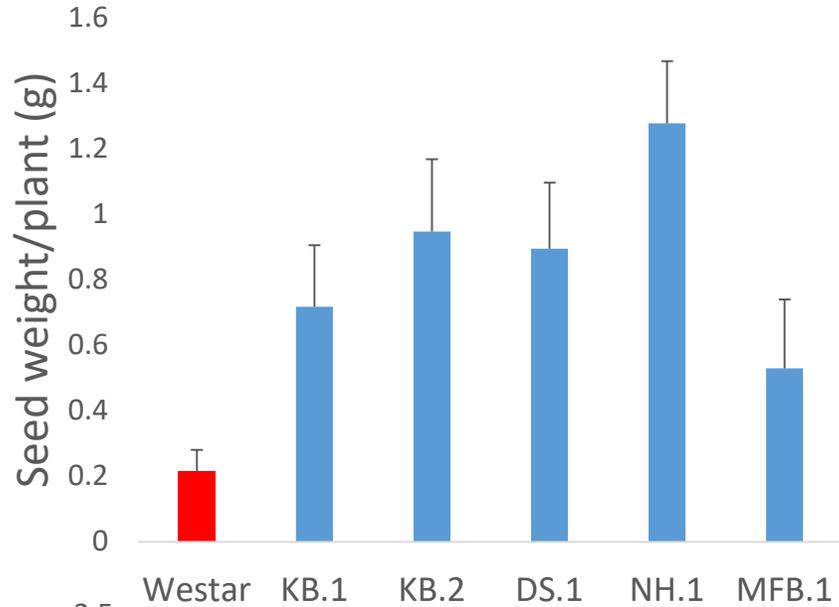


Plants were then removed from chamber and infection continued (1 week)



Control plants dying; transgenic plants producing seed

Does increased protection correlate to increased seed production?



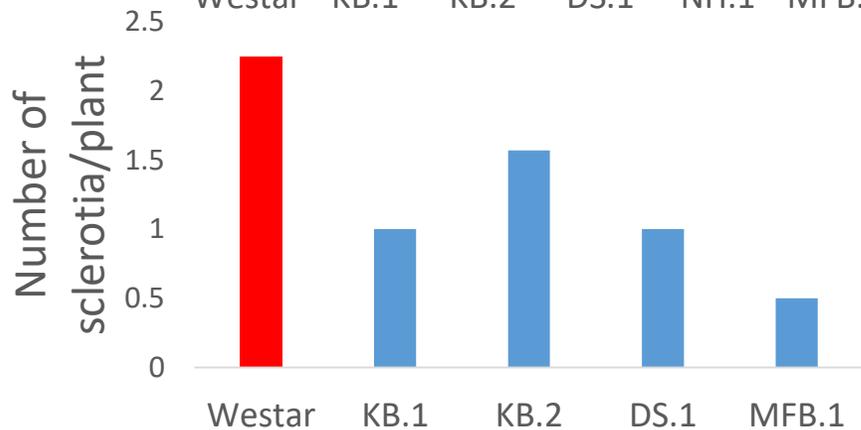
Seed



Westar



Transgenic - RNAi



Sclerotia



On-going activities

- Testing foliar formulations to improve field durability (penetrants and adherents)
- Assessing degradation of dsRNA in soil
- Testing off-target effects
- Field tests of the best performing foliar treatments



The main points

- RNAi offers species-specific control of pest insects and fungal pathogens
- Foliar sprays of dsRNA and transgenic plants are both effective at controlling flea beetles and Sclerotinia
- Field trials will provide conclusive evidence of efficacy and affordability of this new technology



Thanks to:

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