

# Development of a New Clubroot Differential Set

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# Background

- **Multiple strains of *P. brassicae* are known to exist**
  - **Differ in their ability to infect different host species, lines or cultivars**
  - **‘Physiologic specialization’ = the occurrence of multiple races or pathotypes**
- **Breeding efforts must be guided by a good understanding of pathogenic diversity in *P. brassicae* populations!**

# Assessments of Pathogenic Diversity

- Strains of a pathogen are identified by their virulence on a *host differential set*

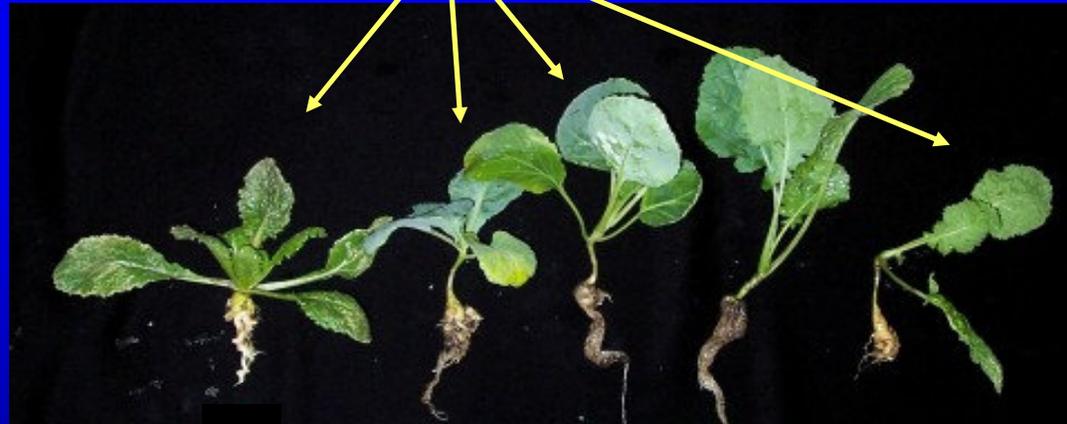
## Differential Set

= A group of host plants that serve to distinguish between various strains of a pathogen based on disease symptoms

(Definition modified from APSnet)



Pathogen



S

S

S

R

S

**Pathogen isolates are grouped into strains based on the symptoms they cause on a defined group of hosts**

# Clubroot Differential Sets

- **Numerous differential sets have been proposed to identify clubroot strains**
- **Three differential sets are most commonly used:**
  - **Williams (1966)**
  - **European Clubroot Differential Set (1975)**
  - **Differentials of Somé et al. (1996)**
- **Each has its advantages & disadvantages**

# Williams' Differential Set

- Developed by P.H. Williams (1966)
- Differential set consists of two rutabagas and two cabbage cultivars
- **Advantage:** Straight-forward and consists of a small set of hosts
- **Disadvantage:** Developed to identify pathogen strains from cabbage and rutabaga

# European Clubroot Differential (ECD) Set

- Developed by Buczacki et al. (1975) as an ‘international system’ for strain identification
- Differential set consists of three subsets:
  - *B. rapa* subset (5 hosts)
  - *B. napus* subset (5 hosts)
  - *B. oleracea* subset (5 hosts)
- **Advantages:** Information on multiple species, enables comparisons
- **Disadvantages:** Lots of hosts, not all hosts differential; complicated strain nomenclature

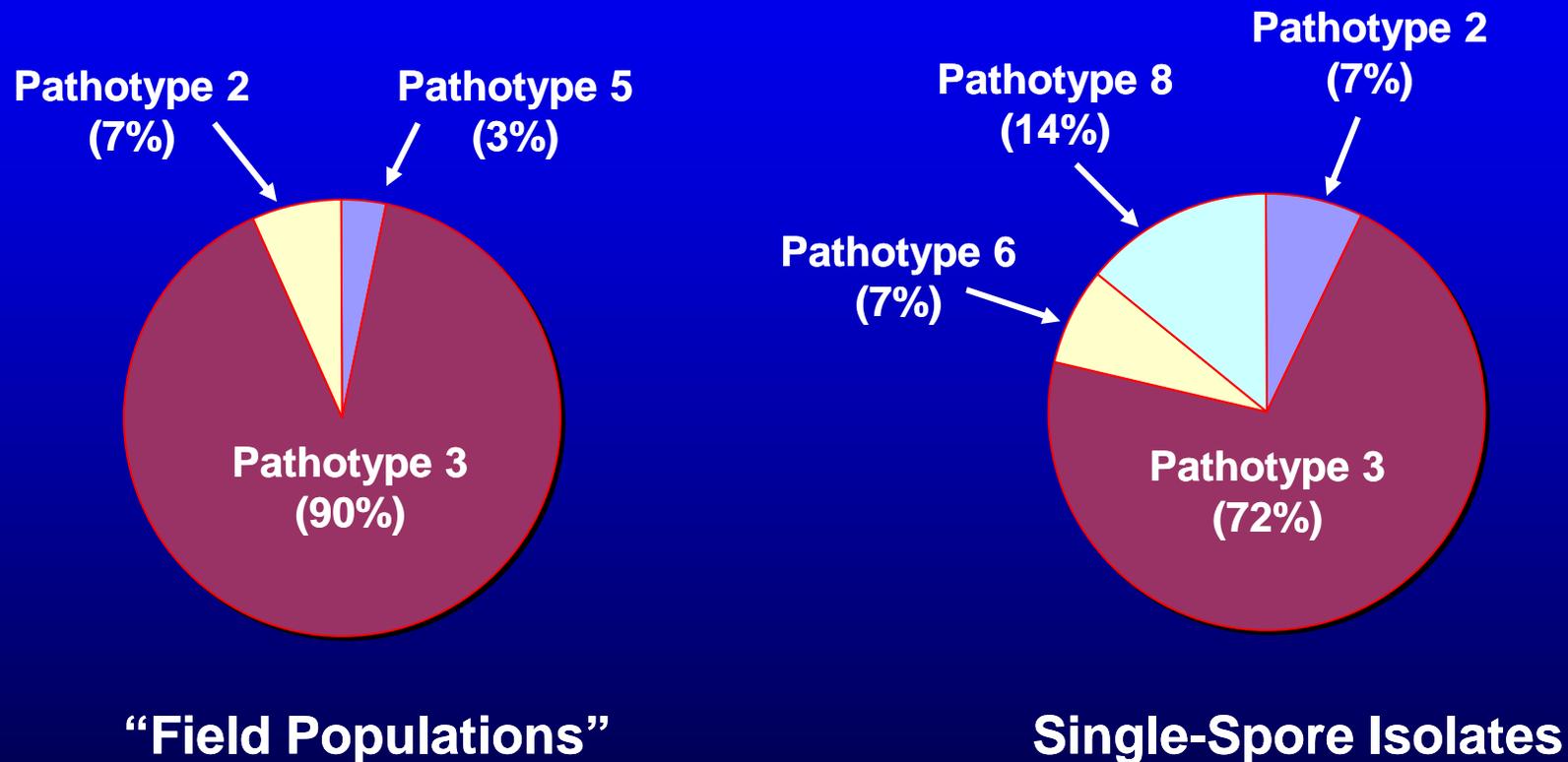
# Differential Set of Somé et al.

- Developed by Somé et al. (1996) to identify pathogen strains from France
- Consists of three *B. napus* hosts
- **Advantages:** Straight-forward and consists of a small set of hosts; based on reaction of *B. napus*
- **Disadvantages:** Low differentiating capacity (we can miss strains)

# Situation in Canada

- **Since the identification of clubroot on canola, we have used all three systems to enable comparisons**
- **Has been effective in identifying predominant strains, but not a perfect system**
- **Challenges:**
  - **Involves a large group of differential hosts**
  - **Some pathotype distinctions relevant for canola, others are not**
  - **May not effectively identify all relevant strains**

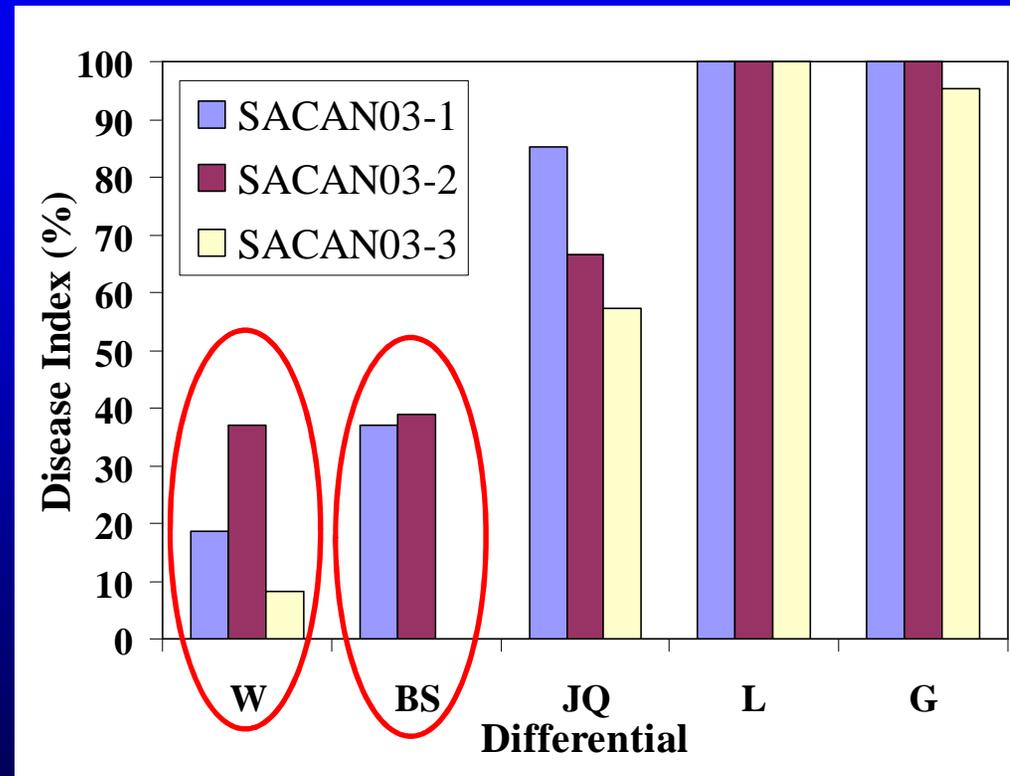
# Strains of *P. brassicae* in Alberta



*Classification on the differentials of Williams (1966)*  
*Pathotype 3  $\approx$  ECD 16/15/12 or  $P_2$  (Some et al. 1996)*

# Strain Identification

- **Another challenge:**
  - **Some differentials give intermediate and fluctuating disease reactions**
  - **What's a resistant reaction and what's not?**
  - **LeBoldus et al. (2012): host considered resistant if index of disease was  $<50\%$  and the 95% CI did not overlap 50%**

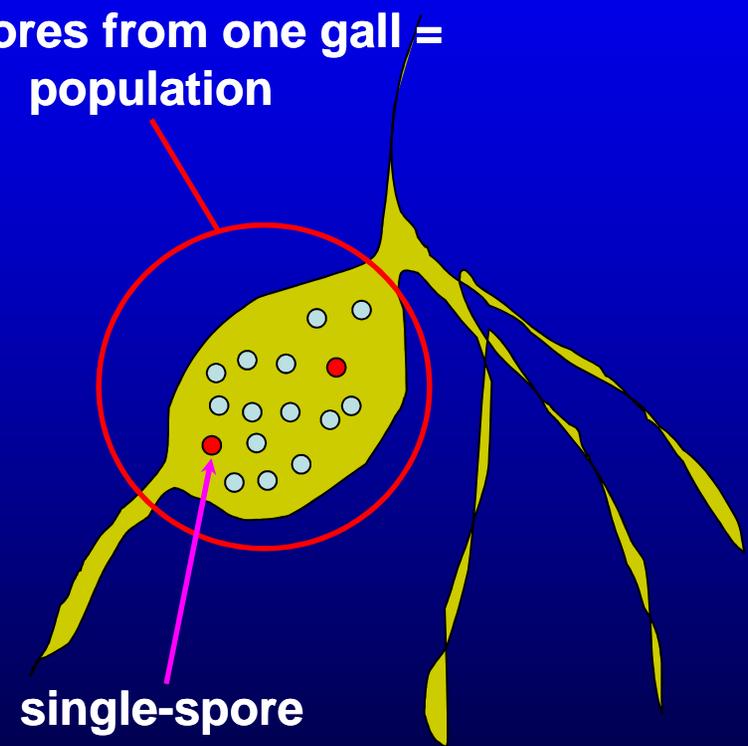


Adapted from Strelkov et al. (2006)

# Fluctuating Reactions

- Largely result of heterogeneity
- In pathogen:
  - Can be addressed by using single-spore isolates instead of populations
- In host:
  - Can be addressed by selecting differentials that give clean reactions

All spores from one gall = population



single-spore

Infected Root

# Pathotypes or Races?

- Largely because of these issues, we refer to clubroot 'pathotypes' instead of 'races'
- Terms are largely synonymous but:
  - 'Pathotype' is a looser term
  - More appropriate because neither the differential hosts nor pathogen populations possess genetic uniformity necessary to apply concept of races to the clubroot pathosystem

# **A New Differential Set?**

- **Given the amount of clubroot work being conducted in Canada and the limitations of existing differentials, a new differential set would be beneficial to identify pathogen strains from canola**

# **Criteria Required of a New Differential Set (According to Strelkov!)**

- **A new set of differentials would have to meet four criteria:**
  - (1) Good differential capacity**
  - (2) Relevance to canola production**
  - (3) Consistent & clear results**
  - (4) Seeds of differentials must be available**

# Development of a New Differential Set

- Using a phased procedure to develop a differential set for *P. brassicae* from canola
  - Consultation of literature & previous studies
  - Screening of *Brassica* genotypes with representative single-spore isolates & populations from Canada
  - Identify subset of putative differentials for screening with wider set of isolates

# Considerations

- Existing differential sets as a starting point
  - Retention of key effective differentials would allow comparisons with literature and international colleagues
- Focus on *B. napus* genotypes with good differentiating capacity, but also include some key *B. rapa* genotypes (exclude *B. oleracea*)
- Include hosts with IDs < 20% or > 80%
  - Avoid hosts with IDs between 20-80% ('indistinct reactions' – Toxopeus et al. 1986)

# *Brassica napus*

- **Greatest differentiating capacity observed in *B. napus* genotypes (both in our tests & in an international analysis)**
  - **Some can distinguish between existing pathotype designations (e.g., pathotype 3 vs. pathotypes 5 & 6)**
  - **Some can differentiate within existing pathotypes (e.g., pathotype 6 isolates from BC & ON)**

Differential Host	Original Pathotype Designation (Differentials of Williams)				
	3	5 (AB)	5 (MB)	6 (BC)	6 (ON)
<b>ECD 06</b>	+	+	+	-	-
<b>ECD 07</b>	+	-	+	+	-
<b>ECD 08</b>	+	+	+	-	-
<b>ECD 09</b>	+	-	+	-	-
<b>ECD 10</b>	-	-	-	-	-
<b>'Brutor'</b>	+	+	+	+	-

*Strelkov, unpublished*

**MB 'pathotype 5' = AB 'pathotype 3'**  
**ON pathotype 6 ≠ BC pathotype 6 (ON strain attacks only cabbage)**

# *Brassica napus*

- **Could also include ‘Mendel’**
- **Some commercial Canadian canola cultivars?**
  - **Two cultivars seem to distinguish pathotype 6 from ON & BC**
  - **Cultivar/germplasm resistant to pathotype 3**
- ***B. napus* susceptible check to replace Chinese cabbage ECD 05?**

# *Brassica rapa*

- ***B. rapa* (Polish rape) hosts ECD 01 – 04 closely related**
  - All are resistant to isolates tested from Canada
  - Also did not contribute to differentiation in an international analysis (Toxopeus et al. 1986)
    - Equally well-represented by ECD 03 alone
- **Worth keeping ECD 02 as resistant check**
  - Prefer ECD 02 to 03 because of clearer reactions in our tests
- **Chinese cabbage (ECD 05) as a susceptible check?**

# Putative Canadian Clubroot Differentials for Further Testing

Common name	Scientific name	Cultivar or line	ECD No.
Polish rape	<i>Brassica rapa</i> var. <i>rapifera</i>	Line AAbbCC	02
Chinese cabbage	<i>B. rapa</i> var. <i>pekinensis</i>	'Granaat'	05
Fodder rape	<i>B. napus</i> var. <i>napus</i>	'Nevin'	06
Fodder rape	<i>B. napus</i> var. <i>napus</i>	'Giant Rape'	07
Fodder rape	<i>B. napus</i> var. <i>napus</i>	Giant Rape Selection	08
Fodder rape	<i>B. napus</i> var. <i>napus</i>	New Zealand Resistant Rape	09
Rutabaga	<i>B. napus</i> var. <i>napobrassica</i>	'Wilhemsburger'	10
Spring oilseed rape	<i>B. napus</i> var. <i>napus</i>	'Brutor'	n/a
Winter oilseed rape	<i>B. napus</i> var. <i>napus</i>	'Mendel'	n/a
Spring canola	<i>B. napus</i> var. <i>napus</i>	'Westar'	n/a
Spring canola	<i>B. napus</i> var. <i>napus</i>	Commercial cv. (R)	n/a
Spring canola	<i>B. napus</i> var. <i>napus</i>	Commercial cv. (S)	n/a

# Advantages of 'Canadian Clubroot Differential' (CCD)

- **Less differential hosts involved**
- **Clearer reactions**
  - If used with single-spore isolates, perhaps could move to a race nomenclature system
- **Better suited to detect variation in pathogenicity on *B. napus* as opposed to cabbage or other hosts**
- **Can compare results obtained with CCD with those obtained with *B. napus* subset of ECD and differentials of Somé et al. (1996)**
  - Facilitate international collaboration & comparisons with historical record

# Next Steps

- **Receive your input!**
- **Inoculate putative differentials with selected single-spore isolates and populations**
- **Finalize list of differentials**
- **Determine race numbering scheme**

# Acknowledgements

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